From gene expression modeling to gene network to investigate Arabidopsis thaliana stress response

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Functional annotation

Definition or prediction of the gene functions and of the relationship between them



- Between 20% and 40% of the predicted genes have no assigned function (Hanson *et al*, 2009)
- For *Arabidopsis thaliana*, only 16% of the genes have a validated function

Orphan genes

- Defined has genes without homologs with a known function (Fukushi and Nishikawa, 2003)
- Usually discarded of the published studies
- 5015 orphan genes in A. thaliana (Zaag et al, 2015)

Based on a comparison of protein sequences to identify structural similarities

Nevertheless

- A high similarity does not guarantee a functional similarity (Tian *et al*, 2003)
- Some sequences with a low similarity may share a same function (Galperin *et al*, 1998)
- Protein sequence comparison gives information about the biochimical function (Nehrt *et al*, 2011)

by omics analysis

Based on guilt by association studies by identification of genes having similar features at the molecular level



Integrating various resources of omics data improves the success of prediction (Radiovojac *et al*, 2013)

But various sources of heterogeneity exist

- Data are qualitative or quantitative
- Available information describes the biological entities or their relationships
- Observations are obtained with various techniques
- Various semantic frameworks are used

From Gene Expression Modeling to Networks



- Required to control the contextual heterogeneity

A dedicated transcriptomic dataset





- 387 transcriptomic comparisons in dye-swap dedicated to stress
- 2/3 describe abiotic stresses and 1/3 biotic stresses
- All the data were generated on the same transcriptomic platform with the same protocol

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First results

- Based on differential analyses, 60% of the genes coding proteins have their transcription impacted directly or not by a stress
- Large overlap of impacted genes between biotic and abiotic stress

Coexpression study using mixture model

what we observe	the model	the expected results	Stress category	Gene_nb	Cluster_nb
			Nitrogen	13 495	59
			Temperature	11 365	34
	ALCONTRACTOR OF		Drought	8 1 4 3	34
7 – ?		Z: 1 = 0, 2 = +, 3 = *	Salt	5 729	30
2 - :			Heavy metal	10 617	57
			UV	7 894	37
Matrix by stress			Gamma	5 3 5 0	32
{ genes x log	g-ratios}	Data-driven method	Oxydative stress	10 127	52
Coursian mir			Nectrophic bacteria	11 220	50
Gaussian mix	ct	number of cluster nosen by BIC	Biotrophic bacteria	12 023	56
		 gene classification based on the conditional 	Fungi	9 773	51
			Rhodococcus	1 900	13
	pi	babilities	Oomycete	5 508	31
*			Nematode	7 413	27
			Stifenia	1 525	17
			Virus	11 832	54
			~ 700 clusters of co-expression		

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Annotation of coexpressed genes



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Visualisation by type of resource



Clustering Biological process Cellular component Molecular function Subcell Bibliostress Orphan Transcription factor Hormone Interactome Network

The GO Biological process was used to characterize the clusters for the stress category VIRUS. Results of gene set enrichment analyses are displayed as one pie chart per cluster, its size reflecting the total number of genes in the cluster.

While the mouse hovers over a pie chart, the total number of genes in cluster appears in a popup and in the 'Biological process' frame on the right side. As well, the number of genes annotated with a GO term is displayed and the hypergeometric test p-value is mentioned when statistical significance is achieved.



Pie size proportional to cluster size Colors indicate biological biases

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Visualisation of interactions



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Overview of a cluster



Overview of all functional annotation analyses made for the cluster 'cluster_49'. While clicking on a circle, a gene set enrichment list for the concerning annotation appears below the main panel. Results of analyses are recapped in the 'Functional Annotation' table on the right side and are downloadable using the link above.



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Vertical integration

Results

- Numerous enrichments
- Overlap with TF regulations and PPI

Conclusions on this large-scale co-expression study

- It generates meaningful groups of genes
- It performs favorably as compared to those obtained with correlation-based approaches (higher % of enrichments)

Nevertheless

- 18 co-expression studies were generated
- Interpretation and use are not straightforward
- Co-expression is not enough to suggest co-regulation and to be used in a guilt by association approach (Dhaeseleer et al., 2000)

Horizontal integration



- Required to control the contextual heterogeneity

From coexpression to coregulation

- Small overlap between two clusters of two different stresses
- Horizontal integration done at the level of the gene pairs



Method

- For each pair of genes, calculation to be in a same cluster of co-expression
- Comparison with a random network: a pair observed more than 3 times is statistically significant (resampling test)

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Coregulation network 5 626 genes and 57 833 interactions

713 orphans and 1 682 with a missing GOSlim annotation



- Degree distribution is a power law
- Considered as an important quality criterion (Gillis et Pavlidis, 2012)



The network with gene pairs conserved in at least 7 stresses 415 genes with 41 orphans, 1 908 interactions



Cis-regulatory motifs found with PLMDetect (Bernard *et al.*, 2010)

- 10 components are enriched in motifs
- For 4 components, the motif is present in over 80% of the gene promotors
- Component 2 has 5 motifs related to the light regulation, present at most in 50% of gene promoters





Conclusions

- Coregulation modules are more specific and more homogeneous
- Cis-regulatory motifs are found in their promoters
- Topological analysis = an approach to identify functional modules

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